

# Interspecies communication in oral biofilm

Esha Tyagi\*, Pragya Jha, Vikas Kacran, Tanushree Bera, Rinky Tripathi

Department of Periodontics, Army College of Dental Sciences, Rajiv Swagruha ABHIMAAN Project, Secunderabad, Telangana 500083  
 Email: [eshashimmer@gmail.com](mailto:eshashimmer@gmail.com)

## Abstract

The diversity of signalling opportunities within microbial communities, and the significant role of these molecules in coordinating gene expression and promoting biofilm formation, has provided the impetus to investigate the potential of inhibitory analogues to disrupt these networks, thereby providing mechanisms to control or influence the development of dental plaque. Within the oral biofilms, resident bacterial cells interact with one another and exchange messages in the form of signalling molecules and metabolites. In this review article, our aim is to elaborate on this mutualistic partnership the role of this quorum sensing and their involvement in pathogenesis to decipher information that can be useful to target pathways to control diseases.

**Key Words:** oral biofilm.

## \*Address for Correspondence:

Dr. Esha Tyagi, Postgraduates, Department of Periodontics, Army College of Dental Sciences, Rajiv Swagruha ABHIMAAN Project, Secunderabad, Telangana 500083

Email: [eshashimmer@gmail.com](mailto:eshashimmer@gmail.com)

Received Date: 23/01/2018 Revised Date: 12/02/2018 Accepted Date: 05/03/2018

DOI: <https://doi.org/10.26611/1019531>

## Access this article online

Quick Response Code:



Website:

[www.medpulse.in](http://www.medpulse.in)

Accessed Date:  
12 March 2018

## INTRODUCTION

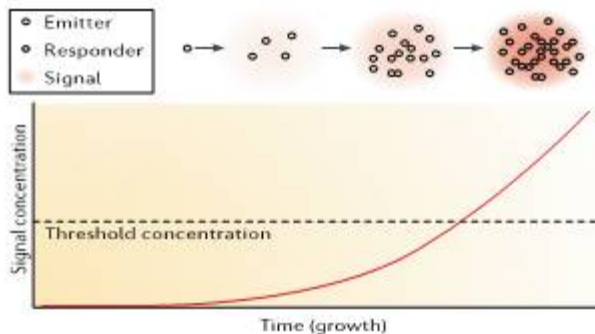
The human fetus inside the uterus is sterile but as soon as it passes through the birth canal, it acquires vaginal and fecal microorganisms. Within 2 weeks, a nearly mature *microbiota* is established in the gut of the newborn baby. Within 2 weeks, a nearly mature *microbiota* is established in the gut of the newborn baby. After weaning (>2 years), the entire human microbiota is formed and comprises a very complex collection of hundreds of different types of

bacteria, totalling approximately  $10^{14}$  microbial cells. It has been estimated that for a normal, healthy human being, the bacterial population comprises 2 kg of the total body weight. The colonization of the oral cavity also starts close to the time of birth. Communication is a key element in successful organizations. The mouth is similar to other habitats within the body in having a characteristic microbial community that provides benefits for the host. The mouth is warm and moist, and is able to support the growth of a distinctive collection of microorganisms (viruses, mycoplasma, bacteria, Archaea, fungi and protozoa). The bacteria on human teeth and oral mucosa have developed the means by which to communicate and thereby form successful organizations. These bacteria have coevolved with their host to establish a highly sophisticated relationship in which both pathogenic and mutualistic bacteria coexist in homeostasis. The foundations of dental plaque are laid by the primary colonizers, predominantly streptococci, actinomyces and a few other genera.

**Table 1:** Properties of biofilms and microbial communities (adapted from Ref.90)

General property	Dental plaque example
Open architecture	Presence of channels and voids
Host protection	Production of extracellular polymers to form a functional matrix; physical protection from phagocytosis
Host protection	Colonization; resistance
Enhanced tolerance to antimicrobials	Reduced sensitivity to chlorhexidine and antibiotics; genetransfer
Neutralization of inhibitors	B-lactamase production by neighbouring cell to protect sensitive organisms
Novel gene expression	Synthesis of novel proteins on attachment or on binding to host molecules; upregulation of gtfBC in mature biofilms
Coordinated gene responses	Production of bacterial cell-to-cell signalling molecules (e.g.) CSP, AI-2)
Communication with host	Downregulation of pro-inflammatory response by resident oral bacteria; remodelling of the cytoskeleton of epithelial cells
Spatial and environment heterogeneity	pH and O <sub>2</sub> gradients; co-adhesion
Broader habitat range	Onligate anaerobes in an overtly aerobic environment
More efficient metabolism	Complete catabolism of complex host macromolecules (e.g. mucins) by microbial consortia (food chains and food webs) pathogenic synergism in periodontal disease

Without retention on the tooth surface, the bacteria are swallowed with the saliva. Through retention, these bacteria can form organized, intimate, multispecies communities referred to as dental plaque. Bacteria have often been studied as populations of cells that act independently, but it now seems that there is much interaction and communication between cells. Bacteria can produce an extensive repertoire of secondary metabolites, and can respond to a wide variety of chemicals in their environment. In recent years, particular groups of secondary metabolites have been characterized for their role in the regulation of gene expression in a cell-density-dependent manner, and this behaviour has been collectively referred to as quorum sensing, or cell-cell communication.



**Figure 1:** scheme for quorum sensing in its simplest form, cell-cells signalling molecules by emitter cell and their accumulation in the signalling molecules bind to receptors on or in the bacterial cell leading to change in gene expression in the responding cell. For intraspecies quorum sensing, the emitter and responder are usually the same cells, as illustrated here. Often, but not always. The genes that are involved in synthesis and response activate their own expression explaining the term autoinducer. A signalling molecule is considered to act at low concentration and not to be involved in primary metabolism.

Dental plaque – a classical multi- species biofilm: Dental plaque has been defined as the microbial community that develops on the tooth surface, embedded in a matrix of polymers of bacterial and salivary origin (90). Dental plaque forms via an ordered sequence of events, resulting in a structurally and functionally organized species-rich microbial biofilm (66, 67, 83, 130). The distinct stages in plaque biofilm formation are described below.

**Reversible adhesion:** Reversible adhesion involves weak, long-range, physico-chemical interactions between the charge on the microbial cell surface and that produced by the conditioning film (8, 19). Microorganisms are usually transported passively to the surface by the flow of saliva or gingival crevicular fluid; a few species (e.g. *Wolinella*, *Selenomonas* and *Campylobacter* spp.) found subgingivally have flagella and are motile.

**Irreversible adhesion:** Irreversible adhesion involves interactions between specific molecules on the microbial cell surface (adhesins) and complementary molecules (receptors) present in the acquired pellicle. These adhesin– receptor interactions are strong and operate over a relatively short distance (159), and are targets for possible novel interventions to block colonization.

**Co-adhesion:** During co-adhesion, secondary and late colonizers adhere here via cell-surface adhesins to receptors on already attached bacteria<sup>65</sup>, leading to an increase in microbial diversity within the developing biofilm (microbial succession) (Fig. 3) (67). Many of the secondary colonizers have fastidious growth requirements. Multiplication of the attached cells leads to an increase in biomass and synthesis of exopolymers to form a biofilm matrix (5, 15). A matrix is a common feature of all biofilms, and is more than a chemical scaffold to maintain the shape of the biofilm. It makes a significant contribution to the structural Plaque as a biofilm and

community – consequences for the microorganisms Dental plaque was the first biofilm to be studied in terms of both its microbial composition and its sensitivity to antimicrobial agents. It is only in recent years, with the advent and application of new molecular and imaging technologies, that a more complete understanding of the biology of dental plaque as a biofilm and microbial community has been possible. Some of the implications of this surface-associated, community-driven lifestyle, and the opportunities for biofilm control, are described below.

### SPATIOTEMPORAL MODEL OF ORAL BACTERIAL COLONIZATION

Development of the oral microbial community involves competition as well as cooperation among the 500 species that compose this community. A few of those oral species are shown in Fig. 1 in a diagram illustrating competition and cooperation among early and late colonizers of the tooth surface. The acquired pellicle, which is composed of a variety of host-derived molecules, coats the enamel surface within minutes after professional cleaning and is a source of receptors recognized by the primary colonizers of dental plaque. These receptors include mucins, agglutinins, proline-rich proteins, phosphate-rich proteins such as statherin, and enzymes such as alpha-amylase. Each is a known receptor for particular oral species.

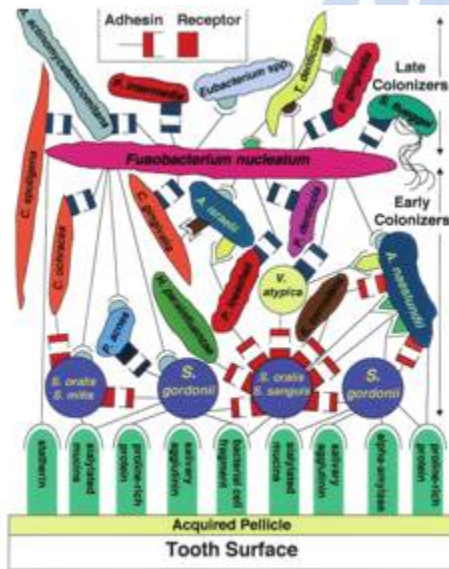


Figure 2:

Streptococci constitute 60 to 90% of the bacteria that colonize the teeth in the first 4 h after professional cleaning (115). Other early colonizers include *Actinomyces* spp., *Capnocytophaga* spp., *Eikenella* spp., *Haemophilus* spp., *Prevotella* spp., *Propionibacterium* spp., and *Veillonella* spp. The complementary symbols

depict physical interactions known to occur between a pair of species. The different shapes and colors of the complementary symbols in represent potentially distinct coaggregations. Rectangle-shaped symbols of any color represent lactose-inhibitable coaggregations, which are prevalent among oral bacteria Those of the same color represent functionally similar but not identical coaggregations. The GalNAc-133 Gal receptor site in 1 Gn is recognized by functionally similar adhesins on *Streptococcus gordonii*, *Haemophilus parainfluenzae*, *Prevotellaloescheii*, *Veillonella atypica*, *Eikenella corrodens*, and *Actinomyces naeslundii*. These adhesins are of various molecular sizes, and the species bearing the adhesins compete with each other for binding to the receptor polysaccharide Thus, it is postulated that coaggregation and coadherence are integral to communication between species and help to establish patterns of spatiotemporal development.

**Metabolic Communication:** The examples of metabolic communication discussed here are limited to interactions in which at least one organism benefits. This arena of metabolic communications among oral bacteria has been reviewed extensively. Beneficial interactions may occur through the excretion of a metabolite by one organism that can be used as a nutrient by a different organism or through the breakdown of a substrate by the extracellular enzymatic activity of one organism that creates biologically available substrates for different organisms. An example of the latter enzymatic activity is sequential hydrolysis of a complex glycoprotein by several bacteria acting in sequence on the product of a previous bacterium's action, as has been shown for oral streptococci Within the oral cavity, bacteria form multispecies communities that are distinguishable primarily by their location. The subgingival community has the highest species richness and the greatest capacity for pathogenic outcome, such as periodontal tissue destruction. In an examination of cocultures of putative periodontal pathogens, such as *P. gingivalis* and *T. denticola*, cocultures produced more biomass than was observed in the respective monocultures; most of the coculture biomass was in the form of cell aggregates, and the coculture was transferable over at least five successive inoculations.

### MECHANISMS OF COMMUNICATION

#### How Bacteria Talk to Each Other: Quorum Sensing

One type of bacterial cell-cell communication is referred to as quorum sensing. Quorum sensing-controlled behaviors are those that only occur when bacteria are at high cell population densities. These behaviors are ones that are unproductive when undertaken by an individual bacterium but become effective by the simultaneous action of a group of cells. For example,

quorum sensing regulates bioluminescence, virulence factor expression, biofilm formation, sporulation, and mating. Quorum sensing is achieved through the production, release, and subsequent detection of and response to threshold concentrations of signal molecules called autoinducers. The accumulation of a stimulatory concentration of an extra- cellular autoinducer can only occur when a sufficient number of cells, a “quorum,” is present. Thus, the pro- cess is proposed to be a mechanism for census taking. There are three archetypal quorum sensing systems (Figure 1; A typical gram-negative bacterial quorum sensing cir- cuit is shown in Figure 1A. In this type of system, the autoinducer is an acylatedhomoserine lactone (AHL) synthesized by a LuxI-type enzyme. Cytoplasmically synthesized autoinducer diffuses passively through the bacterial membrane and accumulates both intra- and extracellularly in proportion to cell density. When the stimulatory concentration of the AHL is achieved, a LuxR-type protein binds it. LuxR-AHL complexes bind to promoters of quorum sensing-regulated target genes and activate transcription. Figure 1B shows the paradigm quorum sensing circuit of a gram-positive bacterium. The autoinducers are short, usually modified peptides processed from precursors. The signals are actively

exported out of the cell, and they interact with the external domains of membrane bound sensor proteins. Signal transduction occurs by a phosphorylation cascade that culminates in the activation of a DNA binding protein that controls transcription of target genes. Specificity exists because each sensor protein is highly selective for a given pep- tide signal. Similar to gram-negative bacteria, gram-pos- itive bacteria can use multiple autoinducers and sen- sors. Some peptide autoinducers act exclusively from the outside, while others elicit a specific set of gene expression changes from the outside and are also trans- ported back into the cell where they trigger a different set of behavioral changes. The final model system shown (Figure 1C) is that of the gram-negative bacterium *Vibrio harveyi*. This quorum sensing circuit controls bioluminescence. *V. harveyi* pro- duces two autoinducers termed HAI-1 and AI-2. HAI-1 is a typical gram-negative-like AHL, although its synthe- sis is not dependent on a LuxI-like enzyme. The second autoinducer, AI-2, is unexpectedly a furanosyl borate diester. HAI-1 and AI-2 signal transduction occurs via a gram-positive-like phosphorylation cascade. Critical for AI-2 signal transduction is the soluble periplasmic AI-2 binding protein LuxP.

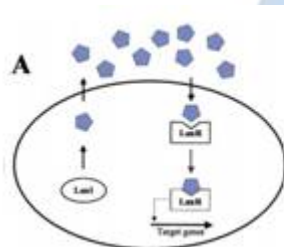


Figure 3:

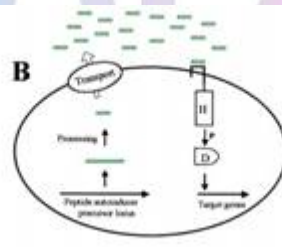


Figure 4:

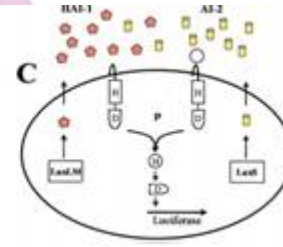


Figure 5:

The tight specificity inherent in AHL and oligopeptide communication circuits presumably results in noise reduction. However, this facet of AHL and oligopeptide circuits also renders them species-specific. In contrast, it appears that the LuxS enzymes from various species of bacteria generate identical intermediates in the AI-2 biosynthetic process. Although at present only one AI-2 structure is known, it is possible that the AI-2s synthesized by different bacteria are identical resulting in a species-nonspecific language.

**Censorship of Free Speech: Anti-Quorum Sensing Strategies** Interspecies cell-cell communication allows bacteria to exploit the diverse metabolic functions that exist in a mixed-species consortium. Several cases of interspecies bacterial communication have been documented, the most notable of which is *V. harveyi* detection of and response to AI-2 produced by

other species of bacteria. AI-2 has only recently been discovered, so its signaling role in these bacteria is not fully clarified. Some bacteria release AI-2 early in growth and internalize at later times. Signal turnover is not a novel feature restricted to AI-2, as it occurs in oligopeptide and AHL signaling systems as well. In these latter cases, elimination of the signal makes possible the initiation and termination of specific behaviors. This could also be the case for AI-2. Alternatively, because AI-2 allows communication between species, bacteria that consume AI-2 could be actively interfering with the signaling process. AI-2 production is integrally associated with SAM metabolism, and therefore, directly tied to cell growth. As such, AI-2 harbors information regarding how well a bacterial population is doing. Elimination of AI-2 from the extra- cellular environment could be a form of “censorship” that allows one species of bacteria to

avoid alerting other species to its presence. Consumption of AI-2 could provide a competitive edge to a bacterial species by rendering other species of bacteria that rely on AI-2 as information at a disadvantage. This could be critical for disrupting the delicate balance that exists between mixed populations competing for colonization of a particular niche. Other examples of anti-quorum sensing “censorship” strategies exist. As mentioned, *S. aureus* groups use peptide quorum sensing to infect a host and to inhibit other *S. aureus* groups from doing so. While the stages of biofilm formation seem to follow basically the same model in various micro-organisms, the biofilm architecture and molecular mechanisms involved in biofilm formation appear to differ.

### CONCLUSION AND FUTURE DIRECTIONS

Developing oral prophylactic strategies through interference with two-component systems or quorum-sensing of biofilm micro-organisms represents an interesting future challenge. Unlike strategies that target microbial viability, such approaches may interfere with microbial adaptive pathways without killing the micro-organisms. Therefore, resistance development would probably represent a minor problem. While the stages of biofilm formation seem to follow basically the same model in various micro-organisms, the biofilm architecture and molecular mechanisms involved in biofilm formation appear to differ. The mechanisms involved in biofilm formation by *P. aeruginosa* are some of the best-characterized and have served as a model for new hypotheses on mechanisms used by other micro-organisms. Information on the genetic regulation of oral biofilm formation, however, is still lacking. A better understanding of these processes is necessary to the development of novel strategies for oral disease prevention and control based on interference of two-component signal transduction systems or quorum-sensing. Since the systems contain both conserved and variable components, both broad- and narrow-spectrum responses may be available. This could allow for tailoring of prophylactic measures based on individual oral health status and risk assessment.

### REFERENCES

1. C. Dobell, “The first observations on entozoic protozoa and bacteria,” in Antony Van Leeuwenhoek and His ‘Little Animals’, pp. 236–256, Russell and Russell, Inc., New York, NY, USA, 1958.
2. P. D. Marsh, “Dental plaque: biological significance of a biofilm and community life-style,” *Journal of Clinical Periodontology*, vol. 32, pp. 7–15, 2005.
3. P. E. Kolenbrander, R. J. Palmer Jr., A. H. Rickard, N. S. Jakubovics, N. I. Chalmers, and P. I. Diaz, “Bacterial interactions and successions during plaque

- development,” *Periodontology 2000*, vol. 42, no. 1, pp. 47–79, 2006.
4. P. E. Kolenbrander, R. N. Andersen, D. S. Blehert, P. G. Eglund, J. S. Foster, and R. J. Palmer Jr., “Communication among oral bacteria,” *Microbiology and Molecular Biology Reviews*, vol. 66, no. 3, pp. 486–505, 2002.
5. R. Huang, M. Li, and R. L. Gregory, “Bacterial interactions in dental biofilm,” *Virulence*, vol. 2, no. 5, pp. 435–444, 2011.
6. J. S. Foster and P. E. Kolenbrander, “Development of a multispecies oral bacterial community in a saliva-conditioned flow cell,” *Applied and Environmental Microbiology*, vol. 70, no. 7, pp. 4340–4348, 2004.
7. H. L. Ritz, “Microbial population shifts in developing human dental plaque,” *Archives of Oral Biology*, vol. 12, no. 12, pp. 1561–1568, 1967.
8. J. Kreth, H. Vu, Y. Zhang, and M. C. Herzberg, “Characterization of hydrogen peroxide-induced DNA release by *Streptococcus sanguinis* and *Streptococcus gordonii*,” *Journal of Bacteriology*, vol. 191, no. 20, pp. 6281–6291, 2009.
9. M. M. Ramsey and M. Whiteley, “Polymicrobial interactions stimulate resistance to host innate immunity through metabolite perception,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 5, pp. 1578–1583, 2009.
10. P. G. Eglund, R. J. Palmer Jr., and P. E. Kolenbrander, “Interspecies communication in *Streptococcus gordonii*-*Veillonellaatypica* biofilms: signaling in flow conditions requires juxtaposition,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 48, pp. 16917–16922, 2004.
11. J. Kreth, J. Merritt, W. Shi, and F. Qi, “Competition and coexistence between *Streptococcus mutans* and *Streptococcus sanguinis* in the dental biofilm,” *Journal of Bacteriology*, vol. 187, no. 21, pp. 7193–7203, 2005.
12. M. Lev, “Sphingolipid biosynthesis and vitamin K metabolism in *Bacteroidesmelaninogenicus*,” *American Journal of Clinical Nutrition*, vol. 32, no. 1, pp. 179–186, 1979.
13. A. P. Roberts and P. Mullany, “Genetic basis of horizontal gene transfer among oral bacteria,” *Periodontology 2000*, vol. 42, no. 1, pp. 36–46, 2006.
14. Y.-H. Li, P. C. Y. Lau, J. H. Lee, R. P. Ellen, and D. G. Cvitkovitch, “Natural genetic transformation of streptococcus mutans growing in biofilms,” *Journal of Bacteriology*, vol. 183, no. 3, pp. 897–908, 2001.
15. K. Willi, H. Sandmeier, S. Asikainen, M. Saarela, and J. Meyer, “Occurrence of temperate bacteriophages in different *Actinobacillusactinomycetemcomitans* serotypes isolated from periodontally healthy individuals,” *Oral Microbiology and Immunology*, vol. 12, no. 1, pp. 40–46, 1997.
16. Signalling molecule involved in bacterial intergeneric communication,” *Microbiology*, vol. 153, no. 10, pp. 3228–3234, 2007. View at Publisher · View at Google Scholar · View at Scopus
17. K. P. Fong, W. O. Chung, R. J. Lamont, and D. R. Demuth, “Intra- and interspecies regulation of gene expression by

- ActinobacillusactinomycetemcomitansLuxS,” *Infection and Immunity*, vol. 69, no. 12, pp. 7625–7634, 2001. View at Publisher · View at Google Scholar · View at Scopus
18. A. H. Rickard, R. J. Palmer Jr., D. S. Blehert et al., “Autoinducer 2: a concentration-dependent signal for mutualistic bacterial biofilm growth,” *Molecular Microbiology*, vol. 60, no. 6, pp. 1446–1456, 2006. View at Publisher · View at Google Scholar · View at Scopus
  19. J. Frias, E. Olle, and M. Alsina, “Periodontal pathogens produce quorum sensing signal molecules,” *Infection and Immunity*, vol. 69, no. 5, pp. 3431–3434, 2001. View at Publisher · View at Google Scholar · View at Scopus
  20. N. S. Jakubovics, “Talk of the town: interspecies communication in oral biofilms,” *Molecular Oral Microbiology*, vol. 25, no. 1, pp. 4–14, 2010. View at Publisher · View at Google Scholar · View at Scopus
  21. J. A. Perry, D. G. Cvitkovitch, and C. M. Lévesque, “Cell death in *Streptococcus mutans* biofilms: a link between CSP and extracellular DNA,” *FEMS Microbiology Letters*, vol. 299, no. 2, pp. 261–266, 2009. View at Publisher · View at Google Scholar · View at Scopus

Source of Support: None Declared  
Conflict of Interest: None Declared

